New acyclic analogues of lipid A: synthesis of 4-phosphonoxybutyl and 3-phosphonoxypropyl glycosides of 2-amino-2-deoxy-D-glucose

Jacques Eustache *, Alfred Grob and Hannelore Retscher Sandoz Forschungsinstitut Wien, Brunner Strasse 59, A-1235 Wien (Austria) (Received January 28th, 1993; accepted June 8th, 1993)

ABSTRACT

Several analogues of lipid A have been synthesized, in which the reducing monosaccharide moiety of the parent molecule has been replaced by an acyclic spacer. The new compounds show high endotoxic activity and are able to protect neutropenic mice against pseudomonas infection, two properties characteristic of LPS-like molecules.

INTRODUCTION

Most of the biological properties of lipopolysaccharides (LPS) have been shown to reside in the lipid A moiety¹. In addition to "beneficial properties" (activation of macrophage/monocytes, B cell mitogenicity, enhancement of host resistance against bacterial or viral infection and tumor)^{1,2}, most lipid A analogues unfortunately show severe toxic side effects, including the well-known and often fatal "endotoxic shock", which severely hampers potential clinical use.

Chemical research in the field of lipid A has developed rapidly following the total synthesis of the *Escherichia coli* lipid A³. A large number of lipid A analogues have since been synthesized and their biological activity examined⁴. Although separation of immunostimulatory and toxic effects could be achieved to a large extent in certain cases, immunostimulatory lipid A-like molecules completely devoid of toxicity have not yet been described. The possibility of designing non-toxic lipid A analogues relies on the hypothesis that different parts of the lipid A molecule are responsible for toxic and immunostimulatory effects. The identification of these structural elements requires systematic chemical modification of the original structure and assessment of corresponding biological activities.

^{*} Corresponding author.

The complexity of the lipid A molecule clearly renders such studies very difficult and therefore our initial aim was to identify a simple structure compatible with high LPS-like activity and which would be amenable to systematic modification. For obvious reasons, in such studies, the molecule used as a starting point should exhibit high biological activity. Examination of the literature suggests that, in the lipid A area, highest biological activities correspond to analogues possessing an intact disaccharide backbone linked to a variable number of suitably located long-chain acyl groups and phosphates⁵. In contrast, monosaccharide analogues are considerably less active. In this work, starting from a disaccharide lead structure we synthesized a series of lipid A analogues in which the reducing sugar unit was replaced by acyclic spacers. We wished primarily to examine the role of the phosphate groups (location and number) with regard to biological activity. The role of the spacer was also studied briefly. Finally, we compared two acylation patterns of the remaining glucosamine unit which had proved to be beneficial with regard to biological activity in the monosaccharide series of lipid A analogues^{4a,6}. These patterns are 2-N-[(R)-3-hydroxytetradecanoyl]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl] and 2-N-[(R)-3-hydroxytetradecanoyl]-3,4-di-O-[(R)-3-hydroxytetradecanoyl].

RESULTS AND DISCUSSION

Synthesis.

2-N-[(R)-3-hydroxytetradecanoyl]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]glucosamine derivatives.—Our synthesis commenced (Scheme 1) with the fully acylated glucosamine derivative 1 which was obtained from the known, 1.3.4,6-tetra-O-acetyl-2-amino-2-deoxy-β-p-glucopyranose⁷, using the mixed anhydride derived from (R)-3-benzyloxymyristoic acid⁸ and isobutyl chloroformate. Compound 1 was glycosylated with 4-allyloxybutanol, using Kiso and Anderson's conditions (FeCl₂, Drierite, CH₂Cl₂), to afford the acetylated glycoside 2. Removal of the ester groups (NaOMe-MeOH) afforded 3 which was tritylated using standard conditions to give 4. Treatment of 4 with (R)-3-tetradecanoyloxytetradecanoic acid¹⁰, 1,3-dicyclohexylcarbodiimide (DCC), and 4-(dimethylamino)pyridine (DMAP) gave 5a and 5b, O-acylated at C-3 and C-4, respectively, which could be separated by column chromatography. The desired key intermediate 5a was isolated, and the minor isomer 5b was deacylated and recycled to afford more 5a; 5a was converted into the desired phosphorylated compound 11 as shown in Schemes 1 and 2. The allyl protecting group was first isomerized [(PPh₃)₃Rh(I)Cl (Wilkinson's catalyst), diazabicyclooctane (DABCO), ethanol] to the corresponding propenyl derivative, obtained as a 1:1 mixture of cis and trans isomers as indicated by proton magnetic resonance (¹H NMR). The propenyl group could be selectively cleaved using aqueous trifluoroacetic acid in THF (under these conditions, the trityl group was unaffected). Selective phosphorylation of the primary hydroxyl group (dibenzyl phosphorochloridate, pyridine, benzene) was followed by removal of the trityl

Scheme 1.

group (HCOOH-ether). Finally, hydrogenolysis of the benzyl groups (10% Pd-C), and addition of one equivalent of tris(hydroxymethyl)aminomethane (TRIS) afforded our first target molecule 11 (Scheme 2).

Using 3-allyloxypropanol instead of 4-allyloxybutanol, the same reaction sequence led to intermediate 8a (Scheme 1), which was converted into the primary phosphate 14 (Schemes 1 and 2).

The 4-phosphate derivative 17 was prepared in 3 steps starting from the intermediate 5a as shown in Scheme 3. Phosphorylation of the free hydroxyl group in 5a (BuLi-dibenzyl phosphorochloridate-THF)¹¹ afforded 15 in moderate yield. First attempts to remove the allyl group in 15, using Wilkinson's catalyst in

Scheme 2.

ethanol, led predominantly to cleavage of the phosphate group. The desired deprotection could be carried out very efficiently using 1,5-cyclooctadienbis(methyldiphenylphosphine)iridium hexafluorophosphate [Ir(COD)PF₆] as a catalyst¹². Thus, the allyl ether 15 was treated under an argon atmosphere with a catalytic amount of the activated catalyst, to afford the corresponding propenyl derivative as a pure *trans* isomer. Hydrolysis of the propenyl group as above using aqueous trifluoroacetic acid in THF followed by trityl cleavage (HCOOH-ether), hydrogenolysis of the benzyl groups, and addition of one equivalent of TRIS afforded 17.

Our first attempts to prepare the protected bisphosphate derivative 18 from 10 were unsuccessful. Due to the low reactivity of the 4-hydroxyl group in 10, harsh phosphorylation conditions (BuLi-dibenzyl phosphorochloridate-THF) are necessary, which are not compatible with the phosphate ester already present in the molecule. Better results were obtained starting from the 4-phosphate derivative 16 and employing milder conditions to introduce the primary phosphate group (dibenzyl phosphorochloridate-pyridine-benzene). Cleavage of the trityl group in 18 (HCOOH-ether), hydrogenolysis of the benzyl groups, and addition of TRIS yielded the bisphosphate derivative 19.

2-N-[(R)-3-hydroxytetradecanoyl]-3,4-di-O-[(R)-3-hydroxytetradecanoyl]glucosamine derivatives. Starting from the trityl derivative 4, the monophosphate 23 (Scheme 4) was prepared. Thus, treatment of 4 with (R)-3-benzyloxytetradecanoic acid and DCC in the presence of DMAP as catalyst gave the triacylated glucosamine derivative 20. Removal of the allyl protecting group [Ir(COD)PF₆] followed by phosphorylation (dibenzyl phosphorochloridate-pyridine-benzene) yielded 22. Cleavage of the trityl ether, hydrogenolysis of the benzyl groups, and addition of TRIS afforded the target molecule 23.

Since the above TRIS salts are not crystalline, slight inaccuracy in the stoichiometry free acid-TRIS is unavoidable. For perfect characterization, analytical

samples of the corresponding free acids were prepared for which correct elemental analyses were obtained in all cases.

Biological activity.

Compounds 11, 14, 17, 19, and 23 were assessed in several assays commonly used for evaluation of lipid A analogues. Compounds 11-19 protected neutropenic mice against pseudomonas infection¹³ and induced endotoxic shock in galactosamine-sensitized mice¹⁴, a characteristic property of LPS-like substances. Compared to our standard 2-deoxy-2-[(R)-3-hydroxytetradecanamido]-3,4-di-O-[(R)-3-hydroxytetradecanoyl]- α -D-glucopyranose 1-phosphate^{4a}, however, the new compounds showed an unfavourable therapeutic ratio. Compound 23, surprisingly, was

only weakly active in the above biological assays. This contrasts with our findings using monosaccharide lipid A analogues, where 2,3,4-N,O-acylated glucosamine derivatives showed a better biological profile^{4a} than 2,3-N,O-acylated derivatives. The effect of the new compounds on B cell activation has been reported elsewhere¹⁵.

The present work constitutes a first step towards the design of structurally simple lipid A analogues. It shows that replacement of one of the monosaccharide units in the lipid A structure can be effected while maintaining biological activity. Based on the present work, further studies aimed at synthesizing even simpler molecules with lipid A-like activity are underway and the corresponding results will be reported in due course¹⁶.

EXPERIMENTAL

General methods.—¹H NMR spectra (internal Me₄Si) were recorded with a Bruker WM-250 (250 MHz) spectrometer. Fast atom bombardment (FAB) mass spectra were recorded with a VG 70-SE spectrometer. Optical rotation measurements were conducted on a Perkin–Elmer 141 polarimeter. Melting points are uncorrected. Thin layer chromatography (TLC) was performed using Silica Gel 60 F₂₅₄ plates (Merck) and column chromatography was done on Merck–Lichroprep columns. Evaporations and concentrations were carried out in vacuo below 40°C unless otherwise specified.

3-Allyloxypropanol and 4-allyloxybutanol.—Sodium hydride (55% in oil; 8.7 g, 200 mmol) was washed 3 times with hexane and suspended in DMF. To the stirred suspension, 1,3-propanediol (30.4 g, 400 mmol) was added slowly, under Ar, while maintaining the temperature below 30°C. Stirring was continued for 2 h and allyl bromide (24.2 g, 200 mmol) was added at such a rate that the temperature did not

In the same way, from 1,4-butanediol (36 g), 4-allyloxybutanol (16.6 g) was obtained; bp 70-75°C (1 mmHg); lit.18 bp 99-104°C (19.5 mmHg). 1H NMR (CDCl₃): δ 1.50–1.90 (4 H, 2 OCH₂CH₂), 2.58 (br s, 1 H, OH), 3.5 (t, 2 H, J 6 Hz, $CH_2CH_2OCH_2CH=CH_2$), 3.66 (br t, 2 H, CH_2CH_2OH), 4.0 (m, 2 H, $OCH_2CH=CH_2$), 5.0–5.50 (2 H, $OCH_2CH=CH_2$), 5.96 (m, 1 H, $OCH_2CH=CH_2$). 1,3,4,6-Tetra-O-acetyl-2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-\(\beta\)-D-glucopyranose (1).—A solution of (R)-3-benzyloxytetradecanoic acid (13.36 g, 40 mmol) and Et₃N (4.04 g, 5.56 mL, 40 mmol) in CH₂Cl₂ (40 mL) was added dropwise to a cold (-30°C) solution of isobutyl chloroformate (5.46 g, 40 mmol) in CH₂Cl₂ (20 mL). The mixture was stirred for 1 h at -30° C and 1 h at 20° C, and a solution of 1,3,4,6-tetra-O-acetyl-2-amino-2-deoxy-\(\beta\)-p-glucopyranose (11.32 g, 32.6 mmol) in CH₂Cl₂ (30 mL) was added dropwise while maintaining the temperature below 25°C. Stirring was continued for 16 h, and the mixture was then concentrated to ca. 50 mL at which point a crystalline precipitate had formed. 1:1 Toluene-hexane (300 mL) was added and the solution was washed with water. The organic layer was dried and evaporated under vacuum, and the residue was dissolved in CH₂Cl₂ and applied to the top of a short column of silica gel (diameter \times height = 10×5 cm). A fast-migrating yellow band was first removed using CH₂Cl₂ as eluant. The desired compound along with slower migrating impurities was eluted using 9:1 CH₂Cl₂-ether. The crystalline residue obtained upon evaporation of the solvent was partially dissolved by refluxing in boiling ether (100 mL) for 1 h. The mixture was stored at 5°C overnight to afford pure 1 as a crystalline powder (15.22 g, 70%); mp 147–148°C; $[\alpha]_D^{20}$ 5.1° (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (t, 3 H, J 6.5 Hz, CH_3), 1.15–1.38 (m, 18 H, 9 alkyl CH_2), 1.40–1.64 (m, 2 H, $BnOCHCH_2CH_2$), 1.99, 2.01, 2.02, 2.07 (4 s, 4×3 H, $4 CH_3CO$), 2.26–2.48 (m, 2 H, BnOCHC H_2CO), 3.66-3.79 (m, 2 H, CHOBn and H-5), 4.12 (dd, $J_{6a,6b}$ 12, $J_{5,6a}$ 2.5 Hz, H-6a), 4.21 (m, 1 H, H-2), 4.29 (dd, $J_{5.6b}$ 4 Hz, H-6b), 4.47 and 4.62 (ABq, 2 H, J_{gem} 12 Hz, $PhCH_2$), 5.03-5.18 (m, 2 H, H-3, 4), 5.58 (d, 1 H, $J_{1,2}$ 8 Hz, H-1), 6.42 (d, 1 H, $J_{1,2}$ 9.5 Hz, NH), 7.30-7.45 (m, 5 H, 5 Ph-H); FABMS: m/z 664 (MH⁺). Anal. Calcd for C₃₅H₅₃NO₁₁: C, 63.33; H, 8.05; N, 2.11; Found: C, 63.47; H, 8.09; N, 2.03.

4-Allyloxybutyl 3,4,6-tri-O-acetyl-2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-β-D-glucopyranoside (2).—A suspension of powdered dry CaSO₄ (13.0 g, 95.5 mmol) in dry CH₂Cl₂ (130 mL) containing FeCl₃ (8.12 g, 50 mmol) and 1 (21.54 g, 32.5 mmol) was stirred for 5 min at 20°C under Ar. 4-Allyloxybutanol (8.45 g, 65 mmol) was added and stirring was continued for 24 h. The mixture was slowly poured into satd aq NaHCO3 and extracted with ether. The organic extract was dried and concentrated under vacuum and the residue thus obtained was chromatographed over silica gel (eluant: 4:1 then 1:1 toluene-EtOAc) to afford pure 2. The compound was dissolved in CH₂Cl₂ and obtained as a crystalline powder (15.6 g, 65%) upon evaporation of the solvent; mp 81.5–85°C; $[\alpha]_D^{20}$ – 5° (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (t, 3 H, J 6.5 Hz, CH₃), 1.15–1.38 (m, 18 H, 9 alkyl CH₂), 1.40–1.70 (m, 6 H, BnOCHC H_2 CH₂ and 2 OCH₂C H_2), 1.97, 2.01, 2.07 (3 s, 3 × 3 H, 3 CH₃CO), 2.30-2.50 (m, 2 H, BnOCHCH₂CO), 3.32-3.45 (m, 3 H, CHOBn and OCH₂CH₂CH₂CH₂O), 3.63 (m, 1 H, H-5), 3.67-3.88 (m, 3 H, H-2 and OCH₂CH₂CH₂CH₂O), 3.94 (m, 2 H, OCH₂CH=CH₂), 4.10 (dd, J_{6a,6b}, 12, J_{5.6a}, 2.5 Hz, H-6a), 4.26 (dd, $J_{5.6b}$ 4 Hz, H-6b), 4.47 (d, 1 H, $J_{1.2}$ 8 Hz, H-1), 4.48 and 4.60 (ABq, 2 H, J_{gem} 12 Hz, PhC H_2), 5.03 (t, $J_{3,4} = J_{4,5} = 10$ Hz, H-4), 5.12-5.32 (3 H, H-3 and OCH₂CH=C H_2), 5.90 (m, 1 H, OCH₂CH=CH₂), 6.51 (d, 1 H, J 8 Hz, NH), 7.28-7.43 (m, 5 H, 5 Ph-H). Anal. Calcd for C₄₀H₆₃NO₁₁: C, 65.46; H, 8.65; N, 1.91; Found: C, 65.56; H, 8.60; N, 1.88.

3-Allyloxypropyl 3,4,6-tri-O-acetyl-2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-β-D-glucopyranoside (6).—From 1 (6.00 g, 9.0 mmol), using the same conditions as for the preparation of 2, 5.26 g (80%) of 6 were obtained; mp 83–85°C; $[\alpha]_D^{20} - 7.3$ ° (*c* 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (t, 3 H, *J* 6.5 Hz, CH₃), 1.15–1.40 (m, 18 H, 9 alkyl CH₂), 1.40–1.70 (m, 2 H, BnOCHC H_2 CH₂), 1.80 (m, 2 H, OCH₂C H_2 CO), 1.98, 2.02, 2.09 (3 s, 3 × 3 H, 3 C H_3 CO), 2.30–2.50 (m, 2 H, BnOCHC H_2 CO), 3.35–3.56 (m, 3 H, CHOBn and OCH₂CH₂C H_2 O), 3.61 (m, 1 H, H-5), 3.68–3.91 (m, 3 H, H-2 and OC H_2 CH₂CH₂O), 3.93 (m, 2 H, OC H_2 CH=CH₂), 4.10 (dd. $I_{6a,6b}$ 12, $I_{5,6a}$ 2.5 Hz, H-6a), 4.26 (dd, $I_{5,6b}$ 4 Hz, H-6b), 4.42 (d, 1 H, $I_{1,2}$ 8 Hz, H-1), 4.47 and 4.60 (ABq, 2 H, I_{gem} 12 Hz, PhC $I_{1,2}$), 5.25 (t, $I_{3,4} = I_{4,5} = 10$ Hz, H-4), 5.12–5.33 (3 H, H-3 and OCH₂CH=C $I_{1,2}$), 5.90 (m, 1 H, OCH₂C $I_{1,2}$ CH=CH₂), 6.50 (d, 1 H, $I_{1,2}$ 8 Hz, NH), 7.25–7.45 (m, 5 H, 5 Ph-H). Anal. Calcd for C₃₉H₆₁NO₁₁: C, 65.07; H, 8.54; N, 1.95; Found: C, 65.20; H, 8.50; N, 1.90.

4-Allyloxybutyl 2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-β-D-glucopyranoside (3).—Compound 2 (10.0 g, 13.6 mmol) was dissolved in dry MeOH (55 mL) and a solution of NaOMe in MeOH (0.1 M, 5.5 mL) was added. The mixture was stirred for 1 h, AcOH (0.033 g, 0.55 mmol) was added, and the MeOH was evaporated. The residue was partitioned between water and CH_2CI_2 , and the organic phase was dried (MgSO₄) and evaporated to dryness to afford 3 as an amorphous solid (8.25 g, 99%); $[\alpha]_D^{20} - 28.9^{\circ}$ (c 1, CHCI₃); ¹H NMR (CDCI₃): δ 0.89 (t, 3 H, J 6.5 Hz, CH₃), 1.20–1.42 (m, 18 H, 9 alkyl CH₂), 1.45–1.70 (m, 6 H, BnOCHC H_2CH_2 and 2 OCH₂C H_2), 2.39 (t, 1 H, J 6.5 Hz, OH), 2.35–2.55 (m, 2 H, BnOCHC H_2CO),

3.12 (br s, 1 H, OH), 3.30–3.65 (7 H), 3.73–3.92 (4 H), 3.94 (m, 2 H, OC H_2 CH=CH $_2$), 4.34 (d, 1 H, $J_{1,2}$ 8 Hz, H-1), 4.52 and 4.57 (ABq, 2 H, J_{gem} 11.5 Hz, PhC H_2), 5.18 (dm, 1 H, J_{cis} 10 Hz, OCH $_2$ CH=C H_{2a} H $_{2b}$), 5.26 (dm, 1 H, J_{trans} 19 Hz, OCH $_2$ CH=CH $_{2a}$ H $_{2b}$), 5.39 (br d, 1 H, J 2.5 Hz, OH), 5.90 (m, 1 H, OCH $_2$ CH=CH $_2$), 6.51 (d, 1 H, J 8 Hz, NH), 7.28–7.43 (m, 5 H, 5 Ph-H). FABMS.: m/z 608 (MH $^+$), 630 (MNa $^+$). Anal. Calcd for C $_{34}$ H $_{57}$ NO $_8$: C, 67.19; H, 9.45; N, 2.30; Found: C, 67.05; H, 9.37; N, 2.29.

4-Allyloxybutyl 2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-trityl-β-D-glucopyranoside (5a).—A mixture of 3 (8.25 g, 13.6 mmol) and trityl chloride (7.25 g, 26 mmol) in pyridine (50 mL) was stirred for 24 h at room temperature. MeOH (11 mL) was then added and stirring was continued overnight. The mixture was partitioned between ether and brine, and the organic phase was washed successively with 1M HCl, satd aq NaHCO₃, and water, dried, and concentrated under reduced pressure. The residue was purified by chromatography under reduced pressure through a short silica gel column (diameter \times height = 9 \times 12 cm). The column was first washed with toluene (1.5 L) which was discarded, then the tritylated compound was eluted using 1:1 EtOActoluene. Evaporation of the solvents afforded the desired trityl derivative 4 (10.9 g, 94%) which was directly used for the subsequent step; ¹H NMR (CDCl₃): δ 0.88 (t, 3 H, J 6.5 Hz, CH₃), 1.18-1.42 (m, 18 H, 9 alkyl CH₂), 1.47-1.73 (m, 6 H, BnOCHC H_2 CH₂ and 2 OCH₂C H_2), 2.35–2.55 (m, 2 H, BnOCHC H_2 CO), 2.78 (br s, 1 H, OH), 3.30-3.65 (7 H), 3.75-3.89 (4 H), 3.92 (m, 2 H, OC H_2 CH=CH₂), 4.34 (d, 1 H, $J_{1.2}$ 8 Hz, H-1), 4.52 and 4.57 (ABq, 2 H, J_{gem} 11.5 Hz, PhC H_2), 5.18 (dm, 1 H, J_{cis} 10 Hz, OCH₂CH=C $H_{2a}H_{2b}$), 5.22 (br s, 1 H, OH), 5.24 (dm, 1 H, J_{trans} 19 Hz, OCH₂CH=CH_{2a} H_{2b}), 5.88 (m, 1 H, OCH₂CH=CH₂), 6.81 (d, 1 H, J8 Hz, NH), 7.15-7.53 (m, 20 H, 20 Ph-H).

A mixture of the trityl derivative 4 (4.37 g, 5.1 mmol), (R)-3-tetradecanoyloxytetradecanoic acid (2.36 g, 5.2 mmol), 1,3-dicyclohexylcarbodiimide (DCC, 1.07 g, 5.2 mmol), and 4-dimethylaminopyridine (DMAP, 0.020 g) in dry CH_2Cl_2 (10 mL) was stirred at 20°C. After 2 h, more acid (0.76 g, 1.68 mmol) and DCC (0.345 g, 1.68 mmol) were added and stirring was continued for 2 h. The precipitated dicyclohexylurea was filtered off, the CH₂Cl₂ evaporated under reduced pressure, and the residue chromatographed over silica gel (eluant: 7:1 toluene-EtOAc) to give 3 fractions from which two compounds homogeneous in TLC (4:1 toluene-EtOAc) were obtained. Fraction I (1.3 g, R_f 0.9), a gum, was not further characterized. Fraction II (3.2 g, R_f 0.56), a gum, was the desired 3-O-acylated compound 5a. Fraction III (1.5 g, R_f 0.47), a gum, was the 4-O-acylated isomer 5b, which was deacylated using the same conditions as for the preparation of 3. Acylation of the recovered 4 (0.81 g) and chromatography as above afforded more **5a**; yield for combined runs: 4.0 g (60%); $[\alpha]_D^{20} - 15.4^\circ$ (c 1, CHCl₃); ¹H NMR $(CDCl_3)$: δ 0.88 (m, 9 H, 3 CH₃), 1.15–1.42 (m, 56 H, 28 alkyl CH₂), 1.42–1.75 (m, 10 H, BnOCHC H_2 CH₂, MyrOCHC H_2 CH₂, C H_2 COO, and 2 OCH₂C H_2), 2.25 (t, 2 H, J 7.5 Hz, CH₂CH₂COO), 2.39 (m, 2 H, BnOCHCH₂CO or MyrOCHC H_2 CO), 2.46 (dd, 1 H, J_{gem} 15, J 4.5 Hz, MyrOCHC H_aH_b CO or BnOCHC H_aH_b CO), 2.57 (dd, 1 H, J 8 Hz, MyrOCHCH $_aH_b$ CO or BnOCHCH $_aH_b$ CO), 3.08 (d, 1 H, J 3 Hz, OH), 3.28–3.47 (6 H), 3.60–4.02 (6 H), 4.17 (d, 1 H, $J_{1,2}$ 8 Hz, H-1), 4.50 and 4.62 (ABq, 2 H, J_{gem} 11.5 Hz, PhC H_2), 4.89 (dd, 1 H, $J_{2,3}$ or 3,4 9, $J_{3,4}$ or 2,3 10.5 Hz, H-3), 5.12 (m, 1 H, MyrOCH), 5.15 (dm, 1 H, J_{cis} 10 Hz, OCH $_2$ CH=C $H_{2a}H_{2b}$), 5.24 (dm, 1 H, J_{trans} 19 Hz, OCH $_2$ CH=CH $_{2a}H_{2b}$), 5.88 (m, 1 H, OCH $_2$ CH=CH $_2$), 6.37 (d, 1 H, J 8 Hz, NH), 7.15–7.53 (m, 20 H, 20 Ph–H). Anal. Calcd for C $_{81}H_{123}$ NO $_{11}$: C, 75.60; H, 9.63; N, 1.09; Found: C, 75.90; H, 10.00; N, 1.20.

3-Allyloxypropyl 2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-6-O-trityl-β-D-glucopyranoside (7).—From **6** (5.39 g), using the same conditions as for the preparation of **3** and **4**, 5.74 g (91%) of 7 were obtained; $[\alpha]_D^{20}$ –31.9° (c 1, CHCl₃); ¹H NMR (1:1 CDCl₃-CD₃OD 1:1): δ 0.89 (t, 3 H, J 6.5 Hz, CH₃), 1.20–1.40 (18 H, 9 alkyl CH₂), 1.45–1.75 (m, 2 H, BnOCHC H_2 CH₂), 1.85 (m, 2 H, OCH₂C H_2 CH₂O), 2.35–2.60 (m, 2 H, BnOCHC H_2 CO), 2.72 (br s, 1 H, OH), 3.30–3,69 (9 H), 3.75–3.79 (4 H), 4.34 (d, 1 H, $J_{1,2}$ 8 Hz, H-1), 4.53 and 4.59 (ABq, 2 H, J_{gem} 11.5 Hz, PhC H_2), 5.15 (dm, 1 H, J_{cis} 10 Hz, OCH₂CH=C H_{2a} H_{2b}), 5.19 (br s, 1 H, OH), 5.24 (dm, 1 H, J_{trans} 19 Hz, OCH₂CH=CH_{2a} H_{2b}), 5.88 (m, 1 H, OCH₂CH=CH₂), 6.76 (d, 1 H, J 8 Hz, NH), (7.15 m, 20 H, 20 Ph-H). Anal. Calcd for C₅₂H₆₉NO₈: C, 74.70; H, 8.32; N, 1.68; Found: C, 74.50; H, 8.10; N, 1.80.

3-Allyloxypropyl-2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-trityl-β-D-glucopyranoside (8a).—From 7 (4.11 9, 4.92 mmol), using the same conditions as for the preparation of 5a, the 3-O-acylated material 8a was obtained [gum, 2.76 g, R_f 0.51 (4:1 toluene-EtOAc)]. The undesired 4-O-acylated isomer 8b (1.32 g, R_f 0.45) was deacylated and reacylated to afford more 8a; yield for combined runs: 3.28 g (52%); $[\alpha]_D^{20}$ -14.2° (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (m, 9 H, 3 CH₃), 1.15–1.42 (m, 56 H, 28 alkyl CH_2), 1.42-1.75 (m, 6 H, BnOCHC H_2 CH₂, MyrOCHC H_2 CH₂, and $CH_2CH_2COO)$, 1.82 (m, 2 H, $OCH_2CH_2CH_2O)$, 2.25 (t, 2 H, J 7.5 Hz, CH_2CH_2COO), 2.39 (m, 2 H, BnOCHC H_2CO or MyrOCHC H_2CO), 2.46 (dd, 1 H, J_{gem} 15, J 4.5 Hz, MyrOCHC H_a H_bCO or BnOCHC H_a H_bCO), 2.57 (dd, 1 H, J8 Hz MyrOCHCH_aH_bCO or BnOCHCH_aH_bCO), 3.08 (d, 1 H, J 3 Hz, OH), 3.28-3.58 (6 H), 3.62-4.02 (6 H), 4.14 (d, 1 H, $J_{1,2}$ 8 Hz, H-1), 4.49 and 4.62 (ABq, 2 H, J_{gem} 11.5 Hz, PhC H_2), 4.87 (dd, 1 H, $J_{2,3 \text{ or } 3,4}$ 9, $J_{3,4 \text{ or } 2,3}$ 10.5 Hz, H-3), 5.12 (m, 1 H, MyrOCH), 5.14 (dm, 1 H, J_{cis} 10 Hz, OCH₂CH=CH_{2a}H_{2b}), 5.23 (dm, 1 H, J_{trans} 19 Hz, OCH₂CH=CH_{2a} H_{2b}), 5.88 (m, 1 H, OCH₂CH=CH₂), 6.33 (d, 1 H, J 8 Hz, NH), 7.15-7.55 (m, 20 H, 20 Ph-H). Anal. Calcd for C₈₀H₁₂₁NO₁₁: C, 75.49; H, 9.58; N, 1.10; Found: C, 75.50; H, 9.20; N, 1.10.

4-Hydroxybutyl 2[(R)-3-benzyloxytetradecanamido]-2-deoxy-3-O-[(R)-3-tetrade-canoyloxytetradecanoyl]-6-O-trityl-β-D glucopyranoside (9).—Compound 5a (4.0 g, 3.1 mmol) was dissolved in EtOH (60 mL), Wilkinson's catalyst (0.63 g, 0.68 mmol) and DABCO (0.216 g, 1.92 mmol) were added, and the mixture was refluxed for 3 h. The solvent was evaporated and the residue was chromatographed over silica gel

(eluant: 9:1 toluene-EtOAc) to afford 4-propenyloxybutyl 2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-trityl- β -D-glucopyranoside (2.93 g). This was dissolved in THF (70 mL) and 9 mL of aq 50% CF₃CO₂H was added. The mixture was stirred for 2 h and more CF₃CO₂H in water (10 mL) was added. After 2 h, the reaction was complete. The mixture was diluted with ether and extracted successively with satd aq NaHCO₃, then water. The organic layer was dried and the solvent evaporated. Chromatography of the residue over silica gel (eluant: 4:1 toluene-EtOAc) afforded pure 9 (1.67 g, 43% from 5a); $[\alpha]_D^{20} - 16.7^{\circ}$ (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (m, 9 H, 3 CH₃), 1.11-1.41 (m, 56 H, 28 alkyl CH_2), 1.41-1.75 (m, 10 H, $BnOCHCH_2CH_2$) MyrOCHC H_2 CH₂, C H_2 CH₂COO, and 2 OCH₂C H_2), 1.92 (br m, 1 H, CH₂OH), 2.25 (t, 2 H, J 7.5 Hz, CH_2CH_2COO), 2.39 (m, 2 H, $BnOCHCH_2CO$ or MyrOCHC H_2 CO), 2.46 (dd, 1 H, J_{gem} 15, J 4.5 Hz, MyrOCHC H_a H_bCO or BnOCHC H_aH_bCO), 2.57 (dd, 1 H, J 8 Hz, MyrOCHC H_aH_bCO or BnOCHCH_aH_bCO), 3.10 (d, 1 H, J 3 Hz, CHOH), 3.28-3.47 (4 H), 3.56 (br m, 2 H, CH₂CH₂OH), 3.70 (m 1 H, H-5), 3.77–3.90 (2 H), 3.98 (m, 1 H, H-2), 4.19 (d, 1 H, $J_{1.2}$ 8 Hz, H-1), 4.52 and 4.62 (ABq, 2 H, J_{gem} 11.5 Hz, PhC H_2) 4.89 (dd, 1 H, $J_{2.3 \text{ or } 3.4}$ 9, $J_{3.4 \text{ or } 2.3}$ 10.5 Hz, H-3), 5.13 (m, 1 H, MyrOCH), 6.31 (d, 1 H, J 8 Hz, NH), 7.15-7.53 (m, 20 H, 20 Ph-H). Anal. Calcd for $C_{78}H_{118}NO_{11}$: C, 75.20; H, 9.55; N, 1.12. Found: C, 75.10; H, 9.20; N, 1.20.

3-Hydroxypropyl 2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-trityl-β-D-glucopyranoside (12).—Using similar conditions as for the preparation of 9, the allyl derivative 8a (3.20 g, 2.51 mmol) was converted into 12 (1.54 g, 50% from 8a); $[\alpha]_D^{20}$ – 16° (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (m, 9 H, 3 CH₃), 1.05–1.42 (m, 56 H, 28 alkyl CH₂), 1.41–1.65 (m, 6 H, BnOCHC H_2 CH₂, MyrOCHC H_2 CH₂, and CH_2 CH₂COO), 1.74 (m, 1 H, OCH₂C H_2 CO), 2.25 (t, 2 H, J 7.5 Hz, CH₂C H_2 COO), 2.39 (m, 3 H, BnOCHC H_2 CO or MyrOCHC H_2 CO + overlapping CH₂OH), 2.46 (dd, 1 H, I_3 Hz, MyrOCHCH_aH_bCO or BnOCHCH_aH_bCO), 3.12 (d, 1 H, I_3 Hz, CHOH), 3.28–3.47 (3 H), 3.53 (m, 1 H), 3.60–3.76 (m, 3H), 3.82 (m, 1 H, CHOBn), 3.88–4.08 (m, 2 H), 4.13 (d, 1 H, $I_{1,2}$ 8 Hz, H-1), 4.51 and 4.64 (ABq, 2 H, $I_{1,2}$ Hz, PhC I_2 CO), 4.84 (dd, 1 H, $I_{2,3}$ or 3,4 9, $I_{3,4}$ or 2,3 10.5 Hz, H-3), 5.13 (m, 1 H, MyrOCH), 6.40 (d, 1 H, I_3 8 Hz, NH), 7.17–7.57 (m, 20 H, 20 Ph-H). Anal. Calcd for $C_{77}H_{117}NO_{11}$: C, 75.02; H, 9.57; N, 1.14. Found: C, 74.52; H, 9.53; N, 1.25.

4-Dibenzyloxyphosphoryloxybutyl 2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-trityl-β-D-glucopyranoside (10).— Compound 9 (0.93 g, 0.75 mmol) was dissolved in dry benzene (10 mL). Pyridine (0.79 g, 10 mmol) and 1 M dibenzyl phosphorochloridate in benzene¹⁹ (2.4 mL) were added. The mixture was stirred for 1.5 h and MeOH (1 mL) was added. Stirring was continued for 10 min, the solvent was evaporated, and the residue was chromatographed over silica gel (eluant: 2:1 toluene–EtOAc) to afford 10 (0.708 g, 63%); $[\alpha]_D^{20} - 13.4^\circ$ (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (m, 9 H, 3 × CH₃),

1.11–1.41 (m, 56 H, 28 alkyl CH₂), 1.41–1.75 (m, 10 H, BnOCHC H_2 CH₂, MyrOCHC H_2 CH₂, C H_2 CH₂COO, and 2 OCH₂C H_2), 2.25 (t, 2 H, J 7.5 Hz, CH₂C H_2 COO), 2.39 (m, 2 H, BnOCHC H_2 CO or MyrOCHC H_2 CO), 2.46 (dd, 1 H, J_{gem} 15, J 4.5 Hz, MyrOCHC H_{a} H_bCO or BnOCHC H_{a} H_bCO), 2.57 (dd, 1 H, J 8 Hz, MyrOCHCH_a H_{b} CO or BnOCHCH_a H_{b} CO), 3.10 (br s, 1 H, OH), 3.26–3.46 (4 H), 3.62–3.83 (3 H), 3.86–4.02 (2 H), 4.16 (d, 1 H, $J_{1,2}$ 8 Hz, H-1), 4.47 and 4.58 (ABq, 2 H, J_{gem} 11.5 Hz, PhC H_2 , 4.90 (dd, 1 H, $J_{2,3 \text{ or } 3,4}$ 9, $J_{3,4 \text{ or } 2,3}$ 10.5 Hz, H-3), 5.00 (d, 4 H, $J_{\text{H,P}}$ 8 Hz, 2 POC H_2 Ph), 5.13 (m, 1 H, MyrOCH), 6.47 (d, 1 H, J 8 Hz, NH), 7.15–7.53 (m, 30 H, 30 Ph-H). Anal. Calcd for C₉₂H₁₃₂NO₁₄P: C, 73.32; H, 8.83; P, 2.06; Found: C, 73.58; H, 8.96; P, 2.18.

3-Dibenzyloxyphosphoryloxypropyl 2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-trityl-β-D-glucopyranoside (13).— Treatment of 12 (0.80 g, 0.65 mmol) as described for the preparation of 10 afforded 13 (0.768 g, 70%); $[\alpha]_D^{20} - 2.8^{\circ}$ (c 0.5, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (m, 9 H, 3 CH₃), 1.11–1.41 (m, 56 H, 28 alkyl CH₂), 1.41–1.75 (m, 8 H, BnOCHC H_2 CH₂, MyrOCHC H_2 CH₂, C H_2 CH₂COO, and OCH₂C H_2 CH₂O), 2.25 (t, 2 H, J 7.5 Hz, CH₂C H_2 COO), 2.30–2.66 (m, 4 H, BnOCHC H_2 CO and MyrOCHC H_2 CO), 3.07 (d, 1 H, J 3 Hz, OH), 3.25–3.45 (4 H), 3.60–3.78 (m, 2 H), 3.80–3.94 (m, 2 H), 4.05 (m, 1 H, H-2), 4.20 (m, 2 H), 4.50 (s, 2 H, PhC H_2), 4.85 (dd, 1 H, $J_{2,3 \text{ or } 3,4}$ 9, $J_{3,4 \text{ or } 2,3}$ 10.5 Hz, H-3), 4.99 (d, 1 H, $J_{H,P}$ 8 Hz, POC H_2 Ph), 5.13 (m, 1 H, MyrOCH), 7.05 (d, 1 H, J 8 Hz, NH),7.15–7.53 (m, 30 H, 30 Ph-H). Anal. Calcd for C₉₁H₁₃₀NO₁₄P: C, 73.21; H, 8.78; P, 2.07; Found: C, 73.10; H, 8.82; P, 2.01.

4-Phosphonoxybutyl 2-deoxy-2-[(R)-3-hydroxytetradecanamido]-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-\(\theta\)-p-glucopyranoside mono TRIS salt (11).—The trityl derivative 10 (0.200 g, 0.13 mmol) was dissolved in ether (20 mL), HCO₂H (16 mL) was added, and the mixture was stirred for 15 min at 20°C. The resulting solution was carefully neutralized by addition of water and solid NaHCO₃, then washed with water, and dried, and the residue was purified by chromatography over silica gel (eluant: 1:1 toluene-EtOAc). The residue (0.147 g) was dissolved in 4:1 THF-water (10 mL). Palladium (10% on charcoal, 0.130 g) was added and the mixture was hydrogenated for 15 min. The catalyst was filtered off over Celite, TRIS (0.015 g, 0.12 mmol) was added, and the solution was lyophilized. In order to eliminate the remaining colloidal particles of carbon, the lyophilizate was dissolved in 8:1 THF-water, and filtered through a short Sephadex LH 20 column (2×5) cm), eluting with the same solvent. The filtrate was lyophilized, and the resulting powder was washed with pentane, filtered, and dried to give pure 11 (0.116 g, 80%). For analytical purposes, a sample of the corresponding free acid 11a was obtained by Bligh-Dyer extraction²⁰: the TRIS salt (0.030 g) was dissolved in a mixture of CHCl₃ (15 mL), MeOH (30 mL), and water (12 mL). Upon addition of CHCl₃ and 0.1 M HCl (15 mL each), two clear phases were obtained. The organic (lower) phase was evaporated to afford 11a; $[\alpha]_D^{20} - 10.4^{\circ}$ (c 0.5, 1:1 CHCl₃-MeOH); ¹H NMR (CDCl₃-CD₃OD + 1 drop of CF₃CO₂D): δ 0.88 (m, 9 H, 3

CH₃), 1.10–1.70 (66 H, 28 alkyl CH₂, HOCHC H_2 CH₂, MyrOCHC H_2 CH₂, C H_2 CH₂COO, and 2 OCH₂C H_2), 2.10–2.31 (4 H, CH₂C H_2 COO and HOCHC H_2 CO or MyrOCHC H_2 CO), 2.45 (dd, 1 H, $J_{\rm gem}$ 15, J 4.5 Hz, MyrOCHC H_a H_bCO or HOCHC H_a H_bCO), 2.56 (dd, 1 H, J 8 Hz, MyrOCHCH_a H_b CO or HOCHCH_a H_b CO), 4.41 (d, 1 H, $J_{1,2}$ 8 Hz, H-1), 4.88 (dd, 1 H, $J_{2,3 \text{ or } 3,4}$ 9, $J_{3,4 \text{ or } 2,3}$ 10.5 Hz, H-3). Anal. Calcd for C₅₂H₁₀₀NO₁₄P: C, 62.81; H, 10.14; P, 3.12; Found: C, 62.31; H, 10.11; P 3.59.

3-Phosphonoxypropyl 2-deoxy-2-[(R)-3-hydroxytetradecanamido]-3-O-[(R)-3-te-tradecanoyloxytetradecanoyl]-β-D-glucopyranoside monoTRIS salt (14).—From the trityl derivative 13 (0.713 g, 0.48 mmol), using the same conditions as for the preparation of 11, the monoTRIS salt 14 (0.415 g, 79%) was obtained. The corresponding free acid 14a was obtained by Bligh-Dyer extraction as described above; $[\alpha]_D^{20} - 14.4^\circ$ (c 0.5, 1:1 CHCl₃-MeOH); ¹H NMR (CDCl₃-CD₃OD 9:1+1 drop of CF₃CO₂D): δ 0.88 (m, 9 H, 3 CH₃), 1.10-1.60 (62 H, 28 alkyl CH₂, HOCHC H_2 CH₂, MyrOCHC H_2 CH₂, and C H_2 CH₂COO), 1.79 (m, 2 H, OCH₂C H_2 CO), 2.08-2.30 (4 H, CH₂C H_2 COO and HOCHC H_2 CO or MyrOCHC H_2 CO), 2.43 (dd, 1 H, J_{gem} 15, J 5 Hz, MyrOCHC H_a H_bCO or HOCHC H_a H_bCO), 2.67 (dd, 1 H, J 7 Hz, MyrOCHCH_aH_bCO or HOCHCH_aH_bCO), 4.39 (d, 1 H, $J_{1,2}$ 8.5 Hz, H-1), 4.83 (dd, 1 H, $J_{2,3}$ or 3,4 9, $J_{3,4}$ or 2,3 10.5 Hz, H-3), 5.04 (m, 1 H, MyrOCH); FABMS: m/z 980 (MH⁺). Anal. Calcd for C₅₁H₉₈NO₁₄P: C, 62.49; H, 10.08; P, 3.16; Found: C, 62.00; H, 10.15; P. 3.69.

4-Allyloxybutyl 2-\(\int(R)\)-3-benzyloxytetradecanamido\(\frac{1}{2}\)-2-deoxy-4-O-dibenzyloxyphosphoryl-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-trityl-β-D-glucopyranoside (15).—To a cold (-78° C) solution of 5a (1.00 g, 0.78 mmol) in THF (80 mL) were successively added a solution of butyllithium (1.6 M in hexane; 0.59 mL, 0.94 mmol) and a 1 M solution of dibenzyl phosphorochloridate in benzene (0.94 mL, 0.94 mmol). After 1 h, AcOH (0.060 g, 1 mmol) was added and the mixture was allowed to warm to room temperature. After evaporation of the solvent, the residue was purified by chromatography over silica gel (eluant: 3:1 toluene-EtOAc) to afford 15 (0.731 g, 61%); $[\alpha]_D^{20} - 9.2^{\circ}$ (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (m, 9 H, 3 CH₃), 1.10-1.80 (66 H, 28 alkyl CH₂, BnOCHCH₂CH₂, MyrOCHC H_2 CH₂, C H_2 CH₂COO, and 2 OCH₂C H_2), 2.25 (t, 2 H, J 7.5 Hz, CH₂CH₂COO), 2.28-2.51 (m, 4 H, BnOCHCH₂CO and MyrOCHCH₂CO), 3.28-3.97 (10 H), 4.32 (ddd, 1 H, $J_{3.4} = J_{4.5} = J_{4.P} = 9$ Hz, H-4), 4.45-4.60 (4 H), 4.60-4.74 (3 H), 5.08-5.27 (3 H, MyrOCH and OCH₂CH=CH₂), 5.28 (dd, 1 H, $J_{2,3 \text{ or } 3,4}$ 9, $J_{3,4 \text{ or } 2,3}$ 10.5 Hz, H-3), 5.88 (m, 1 H, OCH₂CH=CH₂), 6.43 (d, 1 H, J 8 Hz, NH), 7.05-7.53 (m, 30 H, 30 Ph-H). Anal. Calcd for C₉₅H₁₃₆NO₁₄P: C, 73.75; H, 8.86; N, 0.91; Found: C, 73.63; H, 9.04; N, 1.23.

4-Hydroxybutyl 2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-4-O-dibenzyloxy-phosphoryl-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-trityl-β-D-glucopyranoside (16).—The allyl derivative 15 (4.22 g, 2.72 mmol) was dissolved in dry THF (18 mL) under Ar. A solution of Ir(COD)PF₆ (0.025 g) in CH₂Cl₂ (2.5 mL) was added.

The catalyst was activated by allowing a stream of H₂ to pass through the mixture for 5 min, excess of H₂ was replaced by Ar, and stirring was continued for 1 h. After evaporation of the solvent, the crude propenyl derivative thus obtained was dissolved in THF (60 mL) and treated for 30 min with 30 mL of aq 50% CF₃CO₂H. The mixture was carefully neutralized with solid NaHCO₃, ether was added, the organic phase was washed with water and dried, and the solvent was evaporated. The residue was chromatographed over silica gel (eluant: 4:1 toluene-EtOAc) to afford 16 (3.46 g, 84%); $[\alpha]_D^{20}$ -11.2° (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (m, 9 H, 3 CH₃), 1.1-1.40 (56 H, 28 alkyl CH₂), 1.40-1.75 (10 H, BnOCHCH₂CH₂, MyrOCHCH₂CH₂, CH₂CH₂COO, and 2 OCH₂CH₂), 1.85 (br s, 1 H, OH), 2.19 (t, 2 H, J 7.5 Hz, CH₂CH₂COO), 2.28-2.50 (m, 4 H, BnOCHC H_2 CO and MyrOCHC H_2 CO), 3.28–4.00 (9 H), 4.32 (ddd, 1 H, $J_{3,4} = J_{4,5}$ $= J_{4-P} = 9$ Hz, H-4), 4.45-4.74 (7 H), 5.12 (m, 1 H, MyrOCH), 5.24 (dd, 1 H, $J_{2.3 \text{ or } 3.4}$ 9, $J_{3.4 \text{ or } 2.3}$ 10.5 Hz, H-3), 6.33 (d, 1 H, J 8 Hz, NH), 7.05-7.50 (m, 30 H, 30 Ph-H); FABMS: m/z 1530 (MNa⁺). Anal. Calcd for $C_{92}H_{132}NO_{14}P$: C, 73.32; H, 8.83; N, 0.93; Found: C, 73.17; H, 8.94; N, 1.39.

4-Hydroxybutyl 2-deoxy-2-[(R)-3-hydroxytetradecanamido]-4-O-phosphono-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-β-D-glucopyranoside (17a).—Compound 16 (0.80 g, 0.53 mmol) was dissolved in ether (24 mL) and treated with HCO₂H (15 mL). After 5 min, the reaction was worked up as described for 12 and the residue chromatographed over silica gel (eluant: EtOAc), the detritylated material thus obtained (0.375 g) was dissolved in 15 mL 4:1 THF-water, and hydrogenated for 10 min over palladium (10% on carbon). TRIS (0.031 g, 0.26 mmol) was added and the mixture was filtered. The filtrate (which still contained particles of charcoal) was lyophilized to afford the crude TRIS salt 17 (0.330 g). Bligh-Dyer extraction as described for 11 afforded 17a (0.260 g, 49%); $[\alpha]_D^{20} - 8.6^{\circ}$ (c 0.5, 1:1 CHCl₃-CH₃OH); ¹H NMR (4:1 CDCl₃-CD₃OD): δ 0.88 (m, 9 H, 3 CH₃), 1.10-1.75 (66 H, 28 alkyl CH₂, HOCHCH₂CH₂, MyrOCHCH₂CH₂, CH₂CH₂COO, and 2 OCH_2CH_2), 2.10-2.40 (m, 4 H, CH_2CH_2COO and $HOCH_2CH_2CO$ MyrOCHC H_2), 2.54 (dd, 1 H, J_{gem} 15, J 4.5 Hz, MyrOCHC H_aH_bCO or $HOCHCH_aH_bCO$), 2.65 (dd, 1 H, J 8 Hz, $MyrOCHCH_aCH_bCO$ HOCHCH_a H_b CO), 3.28–4.00 (9 H), 4.32 (ddd, 1 H, $J_{3,4} = J_{4,5} = J_{4-P} = 9$ Hz, H-4), 4.54 (d, 1 H, $J_{1.2}$ 8 Hz, H-1), 5.14 (m, 2 H, MyrOCH and H-3). Anal. Calcd for C₅₂H₁₀₀NO₁₄P: C, 62.81; H, 10.14; P, 3.12; Found: C, 62.82; H, 10.24; P. 3.36.

4-Dibenzyloxyphosphoryloxybutyl 2-[(R)-3-benzyloxytetradecanamido]-2-deoxy-2-4-O-dibenzyloxyphosphoryl-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-6-O-trityl-β-D-glucopyranoside (18).—Compound 16 (1.488 g, 0.98 mmol) was dissolved in benzene (20 mL). Pyridine (1.17 mL) and 1 M dibenzyl phosphorochloridate in benzene (4 mL) were added. The mixture was stirred for 3 h at room temperature and MeOH (3 mL) was added. Stirring was continued for 30 min, the solvent was evaporated, and the residue was chromatographed over silica gel (eluant: 3:1 toluene–EtOAc) to afford 18 (1.13 g, 65%); $[\alpha]_D^{20}$ – 7.4° (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (m, 9 H, 3 CH₃), 1.10–1.80 (66 H, 28 alkyl CH₂, BnOCHC H_2 CH₂,

MyrOCHC H_2 CH $_2$, C H_2 CH $_2$ COO, and 2 OCH $_2$ CH $_2$), 2.18 (t, 2 H, J 7.5 Hz, CH $_2$ CH $_2$ COO), 2.28–2.50 (m, 4 H, BnOCHC H_2 CO and MyrOCHC H_2 CO), 3.25–4.05 (9 H), 4.32 (ddd, 1 H, $J_{3,4} = J_{4,5} = J_{4,P} = 9$ Hz, H-4), 4.45–4.75 (7 H), 5.0 (2 overlapping m, 4 H, 2 POC H_2 Ph), 5.14 (m, 1 H, MyrOCH), 5.28 (dd, 1 H, $J_{2,3 \text{ or } 3,4}$ 9, $J_{3,4 \text{ or } 2,3}$ 10.5 Hz, H-3), 6.53 (d, 1 H, J 8 Hz, NH), 7.05–7.50 (m, 40 H, 40 Ph-H). Anal. Calcd for C $_{106}$ H $_{145}$ NO $_{17}$ P $_2$: C, 72.04; H, 8.27; N, 0.79; Found: C, 71.45; H, 8.10; N, 0.77.

4-Phosphonoxybutyl 2-deoxy-2-[(R)-3-hydroxytetradecanamido]-4-O-phosphono-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-\(\beta\)-glucopyranoside bisTRIS salt (19).— The trityl derivative 18 (1.146 g, 0.65 mmol) dissolved in ether (34 mL) was treated with HCO₂H (20 mL) for 5 min. After workup as described earlier, the detritylated material was purified by chromatography over silica gel (eluant: 2:1 toluene-EtOAc) (yield: 0.830 g). The product (0.780 g, 0.51 mmol) was dissolved in 4:1 water-THF (30 mL), and hydrogenated for 30 min over palladium (5% over charcoal, 0.50 g). The catalyst was filtered off, and TRIS (0.121 g, 1 mmol) was added. The solution was lyophilized, and the resulting powder was washed with ether and dried to afford 19 (0.511 g, 64% from 18). An analytical sample of the corresponding free acid 19a was prepared by dissolving 90 mg of the TRIS salt in 15:30:12 CHCl₃-MeOH-water (57 mL), and subjected to Bligh-Dyer extraction as described for 17; $[\alpha]_D^{20}$ 0° (c 0.5, MeOH); ¹H NMR (1:1 CDCl₃-CD₃OD): δ 0.89 (m, 9 H, 3 CH_3), 1.15–1.81 (66 H, 28 alkyl CH_2 , $HOCHCH_2CH_2$, MyrOCHC H_2 CH₂, C H_2 CH₂COO, and 2 OCH₂C H_2), 2.15–2.40 (m, 4 H, CH₂CH₂COO and HOCH₂CH₂CO or MyrOCHCH₂), 2.57 (dd, 1 H, J_{gem} 16, J 6 Hz, MyrOCHC H_a H $_b$ CO or HOCHC H_a H $_b$ CO), 2.69 (dd, 1 H, J 7.5 Hz, MyrOCHCH_aC H_b CO or HOCHCH_a H_b CO), 3.28–3.75 (9 H), 4.28 (ddd, 1 H, $J_{3,4} = J_{4,5} = J_{4,P} = 9$ Hz, H-4), 4.56 (d, 1 H, $J_{1,2}$ 8 Hz, H-1), 5.20 (m, 2 H, MyrOCH and H-3). Anal. Calcd for C₅₂H₁₀₁NO₁₇P₂: C, 58.14; H, 9.48; P, 5.77 Found: C, 57.90; H, 9.36; P, 5.74.

4-Allyloxybutyl 2-[(R)-3-benzyloxytetradecanamido]-3,4-di-O-[(R)-3-benzyloxytetradecanoyl]-2-deoxy-6-O-trityl-β-D-glucopyranoside (20).—Compound 4 (2.92 g, 3.43 mmol) was dissolved in CH₂Cl₂ (10 mL). The solution was cooled to 0°C, and (R)-3-benzyloxytetradecanoic acid (2.34 g, 7 mmol), DCC (1.44 g, 7 mmol), and DMAP (0.020 g) were added. The mixture was stirred for 2.5 h at 0°C, the dicyclohexylurea filtered off, the solvent evaporated, and the residue chromatographed over silica gel (eluant: 9:1 toluene–EtOAc) to afford 20 (4.26 g, 84%); $[\alpha]_D^{20} - 10.5^{\circ}$ (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (t, 9 H, J 6.5 Hz, 3 CH₃), 1.10–1.75 (m, 64 H, 27 alkyl CH₂, 3 BnOCHC H_2 CH₂, and 2 OCH₂C H_2), 2.02 (dd, 1 H) and 2.15–2.50 (m, 5 H) (3 BnOCHC H_2 CO), 3.14 (m, 2 H), 3.35–3.90 (9 H), 3.92 (m, 2 H, OC H_2 CH=CH₂), 4.22–4.62 (m, 7 H, 3 PhC H_2 and H-1), 5.00–5.33 (m, 4 H, OCH₂CH=CH₂), 4.22–4.62 (m, 7 H, 3 PhC H_2 and H-1), 6.22 (d, 1 H, J 8 Hz, NH), 7.15–7.50 (m, 30 H, 30 Ph-H). Anal. Calcd for C₉₅H₁₃₅NO₁₂: C, 76.94; H, 9.17; N, 0.94; Found: C, 76.30; H, 9.21; N, 1.15.

4-Hydroxybutyl 2-[(R)-3-benzyloxytetradecanamido]-3,4-di-O-[(R)-3-benzyloxy-

tetradecanovl 2-deoxy-6-O-trityl-\(\text{B-D-glucopyranoside}\) (21).—The allyl derivative 20 (1.66 g, 1.12 mmol) was dissolved in THF (freshly distilled over LiAlH₄, 7 mL), Ar was bubbled through the solution for 5 min, and Ir(COD)PF₆ (0.008 g) dissolved in CH₂Cl₂ (0.8 mL) was added. Ar was again bubbled for 5 min through the resulting solution and the flask was flushed with H₂ for 1 min under vigorous stirring. Excess of H₂ was removed using a stream of Ar and the mixture was stirred for 30 min. The solvent was evaporated, the residue was dissolved in THF (30 mL), aq 50% CF₃CO₂H (12 mL) was added, and the mixture was stirred for 3 h. Workup as described for 16 and chromatography of the residue over silica gel (eluant: 4:1 toluene-EtOAc) afforded pure 21 (1.37 g, 85%); $[\alpha]_D^{20}$ 6.6° (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (t, 9 H, J 6.5 Hz, 3 CH₃), 1.10-1.75 (m, 64 H, 27 alkyl CH₂, 3 BnOCHC H_2 CH₂, and 2 OCH₂C H_2), 1.85 (br s, 1 H, OH), 2.02 (dd, 1 H) and 2.15-2.50 (m, 5 H) (3 BnOCHCH₂CO), 3.15 (m, 2 H), 3.4-3.97 (9 H), 4.22-4.62 (m, 7 H, 3 PhC H_2 and H-1), 5.08 (dd, 1 H, $J_{3,4} = J_{4,5} = 9$ Hz, H-4), 5.21 (dd, 1 H, $J_{2,3} = J_{3,4} = 10$ Hz, H-3), 6.20 (d, 1 H, J 8 Hz, NH), 7.15–7.50 (m, 30 H, 30 Ph-H). Anal. Calcd for C₉₂H₁₃₁NO₁₂: C, 76.57; H, 9.15; N, 0.97; Found: C, 76.23; H, 9.22; N, 1.18.

4-Dibenzyloxyphosphoryloxybutyl 2-[(R)-3-benzyloxytetradecanamido]-3,4-di-O-[(R)-3-benzyloxytetradecanoyl]-2-deoxy-6-O-trityl-β-D-glucopyranoside (22).—The intermediate 21 (1.26 g, 0.87 mmol) dissolved in benzene (3 mL) was treated for 4 h, at room temperature, with dibenzyl phosphorochloridate (3.8 mL of a 1 M solution in benzene) in the presence of pyridine (1.75 g, 22 mmol) to afford, after purification by chromatography (silica gel, eluant: 6:1 toluene–EtOAc), 22 (0.81 g, 54%); $[\alpha]_D^{20}$ 8.2° (c 0.5, CHCl₃); ¹H NMR (CDCl₃): δ 0.88 (t, 3 H, J 6.5 Hz, CH₃), 1.10–1.75 (m, 64 H, 27 alkyl CH₂, 3 BnOCHCH₂CH₂, and 2 OCH₂CH₂), 2.02 (dd, 1 H) and 2.15–2.50 (m, 5 H) (3 BnOCHCH₂CO), 3.14 (m, 2 H), 3.3–3.05 (9 H), 4.20–4.60 (m, 7 H, 3 PhCH₂ and H-1), 5.08 (dd, 1 H, $J_{3,4} = J_{4,5} = 9$ Hz, H-4), 5.0 (m, 4 H, 2 POCH₂Ph), 5.07 (dd, 1 H $J_{3,4} = J_{4,5} = 9$ Hz, H-4), 5.26 (dd, 1 H, $J_{2,3} = J_{3,4} = 10$ Hz, H-3), 6.37 (d, 1 H, J 8 Hz, NH), 7.15–7.50 (m, 40 H, 40 Ph-H). Anal. Calcd for C₁₀₆H₁₄₄NO₁₅P: C, 74.75; H, 8.52; N, 0.82; Found: C, 74.51; H, 8.60; N, 0.89.

4-Phosphonoxybutyl 2-deoxy-2-[(R)-3-hydroxytetradecanamido]-3,4-di-O-[(R)-3-hydroxytetradecanoyl]-β-D-glucopyranoside monoTRIS salt. (23).—Compound 22 (0.876 g, 0.51 mmol) dissolved in ether (25 mL) was treated with 15 mL of HCO₂H. After 1 h, the reaction was worked up as described previously. Chromatography (3:2 toluene-EtOAc) afforded a syrup (0.589 g) which was dissolved in 4:1 THF-water (25 mL), and hydrogenated for 25 min over 10% palladium-on-charcoal (0.370 g). TRIS (0.049 g, 0.40 mmol) was added and the solution thus obtained was lyophilized. After filtration through a short Sephadex LH20 column and lyophilization, the residue was washed with ether to afford pure 23 (0.352 g, 61% from 22). A small sample of the corresponding free acid 23a was prepared by Bligh-Dyer extraction as described earlier; $[\alpha]_D^{20} - 17^{\circ}$ (c 0.5, 9:1 CHCl₃-MeOH); ¹H NMR (9:1 CDCl₃-CD₃OD + 1 drop of CF₃CO₂D): δ 0.88 (t, 9 H, J 6.5 Hz, 3

CH₃), 1.2–1.8 (m, 64 H, 27 alkyl CH₂, 3 OCHC H_2 CH₂, and 2 OCH₂C H_2), 2.4–2.6 (m, 6 H, 3 HOCHC H_2 CO), 4.62 (d, 1 H, J 8 Hz, H-1), 5.0 (dd, 1 H, $J_{3,4} = J_{4,5} = 9$ Hz, H-4), 5.59 (dd, 1 H, $J_{2,3} = J_{3,4} = 10$ Hz, H-3). Anal. Calcd for C₅₂H₁₀₀NO₁₅P: C, 61.82; H, 9.98; P, 3.07 Found: C, 61.84; H, 10.02; P, 3.19.

ACKNOWLEDGMENTS

We thank Professor J.H. van Boom (Leiden University, The Netherlands) for stimulating discussions and a gift of Ir(COD)PF₆, and Dr G. Schulz of our analytical department for providing and discussing NMR data.

REFERENCES

- O. Lüderitz, C. Galanos, V. Lehman, H. Mayer, E.T. Rietschel, and J. Weckesser, Naturwissenschaften, 65 (1978) 578-585.
- 2 M. Prant, in A. Nowotony (Ed.), Beneficial Effects of Endotoxins, Plenum, New York, 1983, pp 381-395.
- 3 M. Imoto, H. Yoshimura, N. Sagakuchio, S. Kusomoto, and T. Shiba, *Tetrahedron Lett.*, 26 (1985) 1545-1548.
- 4 (a) P.L. Stuetz, H. Aschauer, J. Hildebrandt, C. Lam, H. Loibner, I. Macher, D. Scholz, E. Schuetze, and H. Vyplel, in A. Nowotny, J.J. Spitzer, and E.J. Ziegler (Eds.), Cellular and Molecular Aspects of Endotoxin Reactions, Elsevier, Amsterdam, 1990, pp. 129-144, and references therein; (b) S. Kotani, H. Takada, I. Takahashi, T. Ogawa, M. Tsujimoto, H. Shimauchi, T. Ikeda, H. Okamura, T. Tamura, K. Harada, S. Tanaka, T. Shiba, S. Kusumoto, and T. Shimamoto, Infect. Immun., 54 (1986) 673-682, and references therein.
- 5 I. Takahashi, S. Kotani, H. Takada, M. Tsujimoto, T. Ogawa, T. Shiba, S. Kusumoto, M. Yamamoto, A. Hasegawa, M. Kiso, M. Nishijima, F. Amano, Y. Akamatsu, K. Harada, S. Tanaka, H. Okamura, and T. Tamura, *Infect. Immun.*, 65 (1987) 57-68.
- 6 M. Matsuura, A. Yamamoto, Y. Kojima, J.Y. Homma, M. Kiso, and A. Hasegawa, J. Biochem. (Tokyo), 98 (1985) 1229-1237.
- 7 M. Bergmann and L. Zervas, Ber., 64 (1931) 975-980.
- 8 M. Imoto, H. Yoshimura, M. Yamamoto, T. Shimamoto, S. Kusomoto, and T. Shiba, Bull. Chem. Soc. Jpn., 60 (1987) 2197-2204.
- 9 M. Kiso and L. Anderson, Carbohydr. Res., 72 (1979) C15-C17.
- 10 M. Inage, H. Chaki, M. Imoto, T. Shimamoto, S. Kusumoto, and T. Shiba, Tetrahedron Lett., 24 (1983) 2011-2014; M. Kiso, S. Tanaka, M. Fujita, Y. Fujishima, Y. Ogawa, H. Ishida, and A. Hasegawa, Carbohydr. Res., 162 (1987) 127-140.
- 11 I. Macher, Carbohydr. Res., 162 (1987) 79-84; M. Inage, H. Chaki, S. Kusumoto, and T. Shiba, Chem. Lett., (1982) 1281-1284.
- 12 J.J. Oltwoort, C.A.A. van Boeckel, J.H. De Koning, and J.H. van Boom, Synthesis, (1981) 305-308.
- 13 J.J. Chase, W. Kubey, M.H. Dulek, C.J. Holmes, M.G. Salit, F.C. Pearson, and E. Ribi, Infect. Immun., 53 (1986) 711-712; M. Parant, in A. Nowotny (Ed.), Beneficial Effects of Endotoxins, Plenum, New York and London, 1983, pp 179-196.
- 14 C. Galanos, M.A. Freudenberg, and W. Reutter, Proc. Natl. Acad. Sci. USA, 76 (1979) 5939-5943.
- 15 T. Pedron, R. Girard, J. Eustache, Murty A.R.C. Bulusu, I. Macher, H. Radzyner-Vyplel, P.L. Stütz, and R. Chaby, Int. Immunol., 4 (1992) 533-540.
- 16 For a similar approach see: Murty A.R.C. Bulusu, P. Waldstaetten, J. Hildebrandt, E. Schütze, and G. Schultz, J. Med. Chem., 35 (1992) 3463-3469.
- 17 F. Nerdel, M. Mamluk, and P. Weyerstahl, Justus Liebigs Ann. Chem., 736 (1970) 75-87.
- 18 T.J. Prosser, J. Am. Chem. Soc., 83 (1961) 1701-1704.
- 19 G.W. Kenner, A.R. Todd, and F.J. Weymouth, J. Chem. Soc., (1952) 3675-3681.
- 20 E.G. Bligh and J.J. Dyer, Can. J. Biochem. Physiol., 37 (1959) 911-918.